

Tetrahedron Letters 43 (2002) 105-110

TETRAHEDRON LETTERS

Synthetic studies on the thiostrepton family of peptide antibiotics: synthesis of the tetrasubstituted dehydropiperidine and piperidine cores

Syuhei Higashibayashi, Kimiko Hashimoto* and Masaya Nakata

Department of Applied Chemistry, Faculty of Science and Technology, Keio University, 3-14-1 Hiyoshi, Kohoku-ku, Yokohama 223-8522, Japan

Received 12 October 2001; revised 29 October 2001; accepted 2 November 2001

Abstract—The tetrasubstituted dehydropiperidine and piperidine cores of the thiostrepton family of peptide antibiotics were synthesized which featured the coupling between the azomethine ylide and the enantiopure sulfinimine, and the subsequent stereoselective reduction of the six-membered imine. © 2001 Elsevier Science Ltd. All rights reserved.

In 1955, thiostrepton was isolated from the culture broth of Streptomyces azureus by the Squibb group.¹ Its structure was elucidated by chemical degradation studies,² X-ray crystallographic analysis,³ and NMR studies.⁴ Other structurally related antibiotics, the siomycins,⁵ the thiopeptins,⁶ and Sch 18640,⁷ were also isolated. The characteristic structure of this thiostrepton family of peptide antibiotics is the bicyclic structure containing a tetrasubstituted dehydropiperidine and/or piperidine moiety, a tetrasubstituted dihydroquinoline moiety, four thiazole moieties, a thiazoline moiety, dehydroamino acid moieties, and a dihydroxyisoleucine moiety (Fig. 1). Because of these complex structural features, synthetic studies on the thiostrepton family of antibiotics have scarcely been attempted.⁸ Most efforts have focused on the syntheses of the much simpler thiopeptide antibiotics.^{9,10} These antibiotics show high activities against Gram-positive bacteria and mycobacteria.^{1,5,6,a,b} In this letter, we describe the enantioselective synthesis of the tetrasubstituted dehydropiperidine and piperidine cores, 1 and 2, of the thiostrepton family of antibiotics.

The retrosynthetic analysis is shown in Scheme 1. The tetrasubstituted dehydropiperidine core 1 would be derived from the tetrasubstituted piperidine core 2 by

dehydrogenation.¹¹ The precursor 3 of 2 would be obtained from the six-membered imine derivative 4 by stereoselective reduction. It is likely that the six-membered imine derivative 4 exists in the equilibrium mixture of the five-membered imine derivative **6a** via the intermediate 5a. We anticipated that the six-membered imine 4 in this equilibrium mixture would be preferentially reduced due to steric hindrance around the imine function in the five-membered imine 6a. It is expected that this stereoselective reduction would be realized by considering the stereoelectronic effect shown in Scheme 1. In order to synthesize the equilibrium mixture of 4-6a, we selected the coupling reaction between the azomethine ylide derived from 7 and the chiral sulfinimine 8 or 9, followed by desulfinylation. This coupling reaction may proceed via the 1,3-dipolar cycloaddition¹² (giving **5b**) or the 1,2-addition reaction (giving **6b**). There is a precedent for the 1,3-dipolar cycloaddition between the azomethine ylides derived from the N-benzylidene α -aminoesters and the chiral sulfinimines.^{13,14} The stereochemical outcome described in the literature^{13a} matches our demand when sulfinimine $\mathbf{8}$ is used. On the other hand, when this reaction proceeds via the 1,2-addition reaction, it is possible to control the C6 stereochemistry in 6b by employing sulfinimine 8 or its epimer 9,¹⁵ however, the C5 stereochemistry in **6b** is unpredictable.

One of the key segments, 2,5-dithiazolyl-1,2-dehydropyrrolidine 7,¹⁶ was prepared from the known pyrrolidine dicarboxylic acid derivatives (Schemes 2 and 3).¹⁷ *cis*-1-Boc-2,5-dicarbethoxypyrrolidine 10^{17} (Scheme 2) was hydrolyzed to the acid, which was converted into the

Keywords: thiostrepton; piperidines; sulfinimines; stereoselective reduction.

^{*} Corresponding author. Present address: Plant Science Center, RIKEN, Laboratory for Biochemical Resources, 2-1 Hirosawa, Wako, Saitama 351-0198, Japan. Tel.: +81-48-467-5893; fax: +81-48-467-5407; e-mail: kimikoh@postman.riken.go.jp



Figure 1.



Scheme 1.









mixed anhydride; and into this solution, gaseous NH₃ was introduced. The resulting amide was treated with Lawesson reagent,¹⁸ giving thioamide **11** in 33% overall yield. Treatment of **11** with ethyl bromopyruvate^{9d} in EtOH afforded **12** and its *trans*-isomer **13** in 61 and 17% yields, respectively. Treatment of **12** with *t*-BuOCl¹⁹ in THF followed by triethylamine provided **7** in 95% yield. In addition, *trans*-1-Boc-2,5-dicarb-ethoxypyrrolidine **14**¹⁷ (Scheme 3) was also transformed to **7** by the same procedure as described in the *cis*-series. From thioamide **15**, obtained from **14** in 19% yield together with the 16% yield of the *cis*-isomer **11**, a comparable yield (each 32%) of **12** and **13** was obtained and then **13** was converted into **7** in 76% yield.

Synthesis of another key segment, the chiral sulfinimine **8** or **9**, and its connection with 7 began with the known **16**,^{9d} which was prepared from L-threonine (Scheme 4).

DIBAL reduction of 16 followed by oxidation with chemical manganese dioxide (CMD)²⁰ afforded aldehyde 17 in 73% yield. Condensation of 17 (1 equiv.) in THF with the Davis sulfinamide²¹ **18** (1 equiv.) or **19** (1 equiv.) in the presence of $LiClO_4$ (8 equiv.) and Et_3N (8 equiv.) provided sulfinimine 8 or 9, respectively.²² Each solution of these sulfinimines was directly used in the subsequent coupling. First, based on the results described in the literature,^{13a} dehydropyrrolidine 7 and sulfinimine 8 were selected for the coupling partners. However, this coupling proceeded via the 1,2-addition reaction (not the 1,3-dipolar cycloaddition); furthermore, the following transformation of the coupling product 20 led to the diastereomer of 21 (vide infra) having the opposite configurations at the C2, C5, and C6 positions.²³ Therefore, sulfinimine 9 was next chosen as the coupling partner. To the above-mentioned solution was added 7 at -25°C. After 1 day, the addition product **6b** and its diastereomer²⁴ were obtained in 71 and 17% yields, respectively. The ¹H NMR signal of NHSO (δ 5.79, J=8.7 Hz) in **6b** supported the fivemembered imine structure; however, the C5 and C6 configurations could not be determined at this stage.

With the addition product **6b** in hand, we turned our attention to the final stage (Scheme 5). After desulfinylation of **6b** with TFA in EtOH, the obtained mixture of **4–6a** was subjected to reduction with NaBH₃CN in AcOH/EtOH to afford piperidine **3** in 52% yield as the sole reduction product. Subsequent protection of the oxazoline amine in **3** with Boc₂O followed by condensa-



Scheme 4.



Scheme 5.

tion with Boc-Ala-OH using 2-chloro-1,3-dimethylimidazolidium hexafluorophosphate (CIP)²⁵ and 1hydroxy-7-azabenzotriazole (HOAt) afforded piperidine 2^{26} in 78% yield. The HMBC spectrum of 2 (from H6 to C2) supported the piperidine skeleton and the NOE experiments of 2 supported the relative configurations of the piperidine ring. The absolute structure of 2 was confirmed by its transformation to 21, which was a degradation product from natural thiopeptin B_a.^{27,28} Finally, dehydrogenation of 2 with *t*-BuOCl¹⁹ and triethylamine gave dehydropiperidine 1^{29} in 95% yield.

In summary, we have synthesized, for the first time, the tetrasubstituted dehydropiperidine and piperidine cores, **1** and **2**, of the thiostrepton family of antibiotics via the coupling reaction of the azomethine ylide derived from 7 and the chiral sulfinimine **9**, followed by stereoselective reduction of the six-membered imine function. We believe that the synthetic route described herein is a convergent and unique one. Synthetic studies of other portions in the thiostrepton family of antibiotics are now in progress.

Acknowledgements

This research was supported by a Grant-in Aid for Scientific Research on Priority Areas (A) 'Exploitation of Multi-Element Cyclic Molecules' from the Ministry of Education, Culture, Sports, Science and Technology, Japan.

References

- (a) Pagano, J. F.; Weinstein, M. J.; Stout, H. A.; Donovick, R. Antibiot. Ann. 1955–1956, 554–559; (b) Vandeputte, J.; Dutcher, J. D. Antibiot. Ann. 1955–1956, 560–561; (c) Steinberg, B. A.; Jambor, W. P.; Suydam, L. O. Antibiot. Ann. 1955–1956, 562–565.
- (a) Kenner, G. W.; Sheppard, R. C.; Stehr, C. E. Tetrahedron Lett. 1960, 23–26; (b) Cross, D. F. W.; Kenner, G. W.; Sheppard, R. C.; Stehr, C. E. J. Chem. Soc. 1963, 2143–2150; (c) Bodanszky, M.; Fried, J.; Sheehan, J. T.; Williams, N. J.; Alicino, J.; Cohen, A. I.; Keeler, B. T.; Birkhimer, C. A. J. Am. Chem. Soc. 1964, 86, 2478–2490; (d) Barton, M. A.; Kenner, G. W.; Sheppard, R. C. J. Chem Soc. (C) 1966, 2115–2119; (e) Bodanszky, M.; Scozzie, J. A.; Muramatsu, I. J. Antibiot. 1970, 23, 9–12.
- Anderson, B.; Hodgkin, D. C.; Viswamitra, M. A. Nature 1970, 225, 233–235.
- 4. (a) Tori, K.; Tokura, K.; Okabe, K.; Ebata, M.; Otsuka, H.; Lukacs, G. *Tetrahedron Lett.* 1976, 185–188; (b) Olesker, A.; Valente, L.; Barata, L.; Lukacs, G.; Hull, W. E.; Tori, K.; Tokura, K.; Okabe, K.; Ebata, M.; Otsuka, H. *J. Chem. Soc., Chem. Commun.* 1978, 577–578; (c) Tori, K.; Tokura, K.; Yoshimura, Y.; Okabe, K.; Otsuka, H.; Inagaki, F.; Miyazawa, T. *J. Antibiot.* 1979, *32*, 1072–1077; (d) Tori, K.; Tokura, K.; Yoshimura, Y.;

Terui, Y.; Okabe, K.; Otsuka, H.; Matsushita, K.; Inagaki, F.; Miyazawa, T. J. Antibiot. **1981**, 34, 124–129; (e) Hensens, O. D.; Albers-Schönberg, G.; Anderson, B. F. J. Antibiot. **1983**, 36, 799–813; (f) Hensens, O. D.; Albers-Schönberg, G. J. Antibiot. **1983**, 36, 814–831; (g) Hensens, O. D.; Albers-Schönberg, G. J. Antibiot. **1983**, 36, 832–845; (h) Mocek, U.; Beale, J. M.; Floss, H. G. J. Antibiot. **1989**, 42, 1649–1652.

- (a) Nishimura, H.; Okamoto, S.; Mayama, M.; Ohtsuka, H.; Nakajima, K.; Tawara, K.; Shimohira, M.; Shimaoka, N. J. Antibiot., Ser. A 1961, 14, 255–263; (b) Ebata, M.; Miyazaki, K.; Otsuka, H. J. Antibiot. 1969, 22, 364–368; (c) Tokura, K.; Tori, K.; Yoshimura, Y.; Okabe, K.; Otsuka, H.; Matsushita, K.; Inagaki, F.; Miyazawa, T. J. Antibiot. 1980, 33, 1563–1567. See also Refs 4a,b,c,d.
- (a) Miyairi, N.; Miyoshi, T.; Aoki, H.; Kohsaka, M.; Ikushima, H.; Kunugita, K.; Sakai, H.; Imanaka, H. J. Antibiot. 1970, 23, 113–119; (b) Miyairi, N.; Miyoshi, T.; Aoki, H.; Kohsaka, M.; Ikushima, H.; Kunugita, K.; Sakai, H.; Imanaka, H. Antimicrob. Ag. Chemother. 1972, 1, 192–196; (c) Muramatsu, I.; Hikawa, E.; Hagitani, A.; Miyairi, N. J. Antibiot. 1972, 25, 537–538; (d) Muramatsu, I.; Motoki, Y.; Aoyama, M.; Suzuki, H. J. Antibiot. 1977, 30, 383–387; (e) Hensens, O. D.; Albers– Schönberg, G. Tetrahedron Lett. 1978, 3649–3652; (f) Motoki, Y.; Muramatsu, I. Pept. Chem. 1979, 13–18 See also Refs 4c,d,f,g.
- Puar, M. S.; Ganguly, A. K.; Afonso, A.; Brambilla, R.; Mangiaracina, P.; Sarre, O.; MacFarlane, R. D. J. Am. Chem. Soc. 1981, 103, 5231–5233.
- 8. Only one report on the synthesis of the *N*-protected 2-(1-amino)propenyl-thiazoline-4-carboxylate derivative (a small moiety of siomycin A and thiopeptin Ba), see: Shin, C.-g.; Ito, A.; Okumura, K.; Nakamura, Y. *Chem. Lett.* **1995**, 45–46.
- Examples of syntheses of micrococcins, see: (a) Kelly, T. R.; Jagoe, C. T.; Gu, Z. Tetrahedron Lett. 1991, 32, 4263–4266; (b) Nakamura, Y.; Shin, C.-g.; Umemura, K.; Yoshimura, J. Chem. Lett. 1992, 1005–1008; (c) Okumura, K.; Shigekuni, M.; Nakamura, Y.; Shin, C.-g. Chem. Lett. 1996, 1025–1026; (d) Ciufolini, M. A.; Shen, Y. C. J. Org. Chem. 1997, 62, 3804–3805; (e) Shin, C.-g.; Okumura, K.; Shigekuni, M.; Nakamura, Y. Chem. Lett. 1998, 139–140; (f) Okumura, K.; Ito, A.; Yoshioka, D.; Shin, C.-g. Heterocycles 1998, 48, 1319–1324; (g) Okumura, K.; Nakamura, Y.; Shin, C.-g. Bull. Chem. Soc. Jpn. 1999, 72, 1561–1569; (h) Okumura, K.; Suzuki, T.; Nakamura, Y.; Shin, C.-g. Bull. Chem. Soc. Jpn. 1999, 72, 2483–2490; (i) Ciufolini, M. A.; Shen, Y.-C. Org. Lett. 1999, 1, 1843–1846.
- An example of the synthesis of promothiocin A, see: (a) Moody, C. J.; Bagley, M. C. *Chem. Commun.* **1998**, 2049–2050; (b) Bagley, M. C.; Bashford, K. E.; Hesketh, C. L.; Moody, C. J. *J. Am. Chem. Soc.* **2000**, *122*, 3301–3313.
- The C2-hydrogen in 2 is more acidic than the C6-hydrogen, see: (a) Meyers, A. I.; Knaus, G. N. J. Am. Chem. Soc. 1973, 95, 3408–3410; (b) Knaus, G.; Meyers, A. I. J. Org. Chem. 1974, 39, 1189–1192; (c) Knaus, G.; Meyers, A. I. J. Org. Chem. 1974, 39, 1192–1195. The numbering system depicted in Scheme 1 is employed for convenience.

- Vivanco, S.; Lecea, B.; Arrieta, A.; Prieto, P.; Morao, I.; Linden, A.; Cossío, F. P. J. Am. Chem. Soc. 2000, 122, 6078–6092 and references cited therein.
- (a) Viso, A.; Fernández de la Pradilla, R.; Guerrero-Strachan, C.; Alonso, M.; Martínez-Ripoll, M.; André, I. J. Org. Chem. 1997, 62, 2316–2317; (b) Viso, A.; Fernández de la Pradilla, R.; García, A.; Alonso, M.; Guerrero-Strachan, C.; Fonseca, I. Synlett 1999, 1543–1546.
- There is also a precedent for the 1,3-dipolar cycloaddition between the azomethine ylide derived from 2,5-dicarbethoxy-1,2-dehydropyrrolidine and *N*-methylmaleimide, see: Husinec, S.; Milovanovic, L.; Savic, V. J. Serb. Chem. Soc. 1996, 61, 523–528.
- Davis, F. A.; Zhou, P.; Chen, B.-C. Chem. Soc. Rev. 1998, 27, 13–18.
- 16. Satisfactory analytical data (NMR and IR spectra, elemental analyses and/or HRMS, optical rotations) were obtained for all new compounds.
- 17. Kemp, D. S.; Curran, T. P. J. Org. Chem. 1988, 53, 5729–5731.
- Thomsen, I.; Clausen, K.; Scheibye, S.; Lawesson, S.-O. In *Organic Syntheses*; Freeman, J. P., Ed.; John Wiley & Sons: New York, 1990; Collect Vol. 7, pp. 372–375.
- (a) Bachmann, W. E.; Cava, M. P.; Dreiding, A. S. J. Am. Chem. Soc. 1954, 76, 5554–5555; (b) Scully, F. E. Jr.; Davis, R. C. J. Org. Chem. 1978, 43, 1467–1468.
- Aoyama, T.; Sonoda, N.; Yamauchi, M.; Toriyama, K.; Anzai, M.; Ando, A.; Shioiri, T. *Synlett* **1998**, 35–36.
- Davis, F. A.; Reddy, R. E.; Szewczyk, J. M.; Reddy, G. V.; Portonovo, P. S.; Zhang, H.; Fanelli, D.; Reddy, R. T.; Zhou, P.; Carroll, P. J. *J. Org. Chem.* **1997**, *62*, 2555–2563.
- In our case, the mixture of LiClO₄-Et₃N,¹² which was very effective for the subsequent one-pot coupling, was convenient for this condensation. For other dehydration reagents, see; (a) Liu, G.; Cogan, D. A.; Ellman, J. A. J. Am. Chem. Soc. 1997, 119, 9913-9914; (b) Liu, G.; Cogan, D. A.; Owens, T. D.; Tang, T. P.; Ellman, J. A. J. Org. Chem. 1999, 64, 1278-1284; (c) Davis, F. A.; Zhang, Y.; Andemichael, Y.; Fang, T.; Fanelli, D. L.; Zhang, H. J. Org. Chem. 1999, 64, 1403-1406.
- 23. The details will be described in a full account.
- 24. The stereochemistry of this diastereomer has not yet been determined.
- (a) Akaji, K.; Kuriyama, N.; Kiso, Y. Tetrahedron Lett. 1994, 35, 3315–3318; (b) Akaji, K.; Kuriyama, N.; Kiso, Y. J. Org. Chem. 1996, 61, 3350–3357.
- 26. Compound **2**: $[\alpha]_{D}^{30}$ +20.9 (*c* 1.00, MeOH); IR (CHCl₃): 1820, 1795, 1720, 1500, 1370, 1325, 1100, 1070, and 1045 cm⁻¹. ¹H NMR (CDCl₃, 300 MHz): δ 8.54 (1H, br s), 8.12 (1H, s), 7.84 (1H, s), 6.89 (1H, s), 5.45 (1H, d, J=6.0 Hz), 5.17 (1H, d, J=2.7 Hz), 5.03 (1H, dq, J=2.7 and 6.3 Hz), 4.59 (1H, s), 4.46 (1H, dd, J=2.7 and 10.1 Hz), 4.41 (2H, q, J=7.2 Hz), 4.40 (2H, q, J=7.2 Hz), 4.16 (1H, dq, J=6.0 and 6.6 Hz), 3.51 (1H, ddd, J=3.0, 3.0,and 14.1 Hz), 2.72 (1H, ddd, J = 3.6, 14.1, and 14.1 Hz), 2.24 (1H, dddd, J=2.7, 3.0, 3.6, and 18.0 Hz), 2.01 (1H, dddd, J=3.0, 10.1, 14.1, and 18.0 Hz), 1.61 (3H, d, J=6.3 Hz), 1.46 (9H, s), 1.45 (3H, d, J=6.6 Hz), 1.39 (6H, t, J=7.2 Hz), 1.30 (9H, s). ¹³C NMR (CDCl₃, 75 MHz): δ 174.9, 173.3, 173.0, 167.4, 161.4, 161.3, 155.3, 152.8, 151.3, 148.8, 146.8, 127.3, 127.1, 119.7, 85.4, 79.7, 75.8, 64.9, 61.5, 61.4, 61.3, 61.2, 58.0, 51.4, 30.7, 28.2,

28.1, 27.8, 27.4, 20.4, 18.7, 14.3. HRMS (FAB⁺) calcd for $C_{37}H_{50}N_7O_{11}S_3$ (M+H)⁺: 864.2731. Found: 864.2726.

27. According to Ref. 6f, the authors have isolated two degradation products, IIA and IIB, from thiopeptin B_a as the piperidine residue. Compound IIA, the epimer of compound **IIB** at the C2 position in the piperidine ring, was formed from IIB as an artifact under their degradation conditions. Therefore, compound IIB was the intact piperidine residue. Unfortunately, they could not analyze the coupling constant $J_{2,3}$ of compound **IIB** because the C2 proton signal overlapped with the C6 proton signal at the 100 MHz NMR spectrometer (see Ref. 28). Instead, judging from the coupling constant $(J_{2,3}=3.5 \text{ and } 12.0 \text{ })$ Hz) in the ¹H NMR spectrum of compound IIA together with X-ray analysis of thiostrepton,³ they proposed that compound IIA was 21 and hence the C2 configuration of the piperidine ring in natural thiopeptin B_a was S. On the other hand, according to Ref. 4f, the authors proposed the C2 configuration of the piperidine ring in thiopeptin **B**_a to be *R* judging from the coupling constant $(J_{2,3}=3.5)$ and 10.0 Hz) in the ¹H NMR spectrum of thiopeptin A_{1a} (thiopeptin A_{1a} is methyl ester of thiopeptin B_a at the terminal position in R⁵ depicted in Fig. 1). These two arguments conflicted. Therefore, we did NOE experiments for both 2 and synthetic 21 that are shown in Scheme 5 and unambiguously determined the C2, C5, and C6 relative configurations. Additionally, the absolute configuration of the piperidine ring in synthetic 21 was confirmed by comparison of its optical rotation with that of the compound IIB described in Refs. 6f and 28. Compound **21**: $[\alpha]_{D}^{30}$ +45.6 (*c* 1.02, 1 M HCl) [lit.^{6f,28} $[\alpha]_{D}^{26}$ +44.7 (c 1.02, 1 M HCl)]. UV (1 M HCl) λ_{max} 239 nm (ε 19500) [lit.^{6f,28} UV (1 M HCl) λ_{max} 240 nm (ε 16900)]. IR (KBr): 1685, 1580, and 1380 cm⁻¹ [lit.^{6f,28} IR (KBr): 1690, 1580, 1480, and 1370 cm⁻¹]. ¹H NMR (D₂O, 300 MHz): δ 8.64 (1H, s), 8.49 (1H, s), 7.59 (1H, s), 5.45 (1H, s), 5.37 (1H, br d, J = 16.5 Hz), 4.94 (1H, d, J = 6.9 Hz), 4.50 (1H, q, J=6.9 Hz),), 4.38 (1H, dq, J=6.6 and 6.9 Hz), 3.44 (1H, br d, J=14.7 Hz), 2.94 (1H, br dd, J = 12.6 and 14.7 Hz), 2.79 (1H, br d, J = 12.6 Hz), 2.47 (1H, br ddd, 12.6, 12.6, and 16.5 Hz), 1.77 (3H, d, J=6.9 Hz), 1.32 (3H, d, J = 6.6 Hz) [lit.²⁸ ¹H NMR (D₂O, 100 MHz): δ 8.61 (1H, s), 8.46 (1H, s), 7.59 (1H, s), 5.48 (1H, s), 5.40 (1H), 5.0-4.7 (1H), 4.52 (1H, q, J=8 Hz), 4.40 (1H, dq, J=6.5 and 6.5 Hz), 3.6-3.3 (1H), 3.2-2.2 (3H),1.77 (3H, d, J=8 Hz), 1.30 (3H, d, J=6.5 Hz)]. ¹³C NMR (D₂O, 75 MHz): δ 172.3, 171.5, 166.4, 166.2 165.2, 164.9, 147.5, 147.2, 146.2, 132.5, 132.1, 126.5, 68.9, 62.6, 62.2, 58.4, 58.3, 51.3, 32.1, 26.3, 20.1, 18.1 [lit.²⁸ ¹³C NMR (D₂O, 25 MHz): δ 172.3, 171.4, 166.4, 165.8 165.2, 164.8, 147.4, 147.1, 146.0, 132.6, 132.1, 126.6, 68.9, 62.6, 62.2, 58.4, 58.4, 51.4, 32.2, 26.2, 20.2, 18.2].

- Motoki, Y. Ph.D. Thesis, Department of Chemistry, College of Science, Rikkyo University, 1980.
- 29. Compound 1: $[\alpha]_D^{30}$ +23.3 (*c* 1.00, CHCl₃). IR (CHCl₃): 1820, 1720, 1500, 1370, 1330, 1160, 1100, 1070, and 1045 cm⁻¹. ¹H NMR (CDCl₃, 300 MHz): δ 8.43 (1H, br s), 8.19 (1H, s), 7.91 (1H, s), 7.00 (1H, s), 5.43 (1H, br s), 5.30–5.12 (3H, m), 4.51–4.34 (4H, m), 3.99 (1H, dq, J=5.7 and 7.2 Hz), 3.63 (1H, ddd, J=0.0, 5.4 and 13.8 Hz), 3.46–3.31 (1H, m), 3.12–2.92 (1H, m), 2.80 (1H, ddd, J=6.0, 13.2, and 13.8 Hz), 1.61 (3H, d, J=6.3 Hz), 1.50 (9H, s), 1.47–1.35 (6H, m), 1.35 (3H, d, J=7.2 Hz), 1.21 (9H, s). ¹³C NMR (CDCl₃, 75 MHz): δ 175.2, 173.7, 168.9, 167.6, 163.6, 161.2, 161.1, 155.2, 153.2, 150.5, 148.6, 147.9, 147.0, 130.0, 127.3, 120.0, 85.2, 79.6, 74.9, 66.7, 61.6, 61.4, 61.3, 59.8, 51.9, 27.9, 27.8, 26.9, 24.5, 20.5, 17.9, 14.2, 14.2. HRMS (FAB⁺) calcd for C₃₇H₄₈N₇O₁₁S₃ (M+H)⁺: 862.2574. Found: 862.2573.